

REFERENCES

- Abdel-Wahab, M. A., El-Samawaty, A. E. R. M., Elgorban, A. M., & Bahkali, A. H. (2021). Thraustochytrids from the Red Sea mangroves in Saudi Arabia and their abilities to produce docosahexaenoic acid. *Botanica Marina*, 64(6), 489-501.
- Al-Khikani, F., & Ayit, A. (2022). The Antibacterial Action of Safranin and Gentian Violet. *Rambam Maimonides Medical Journal*, 13(3).
- Arifanti, V. B., Sidik, F., Mulyanto, B., Susilowati, A., Wahyuni, T., Yuniarti, N., ... & Novita, N. (2022). Challenges and strategies for sustainable mangrove management in Indonesia: a review. *Forests*, 13(5), 695.
- Bagul, V. P., & Annapure, U. S. (2021). Isolation and characterization of docosahexaenoic acid-producing novel strain *Aurantiochytrium* sp. ICTFD5: A sterol with vitamin d-cholecalciferol, and cellulase and lipase producing thraustochytrid. *Bioresource Technology Reports*, 14, 100688.
- Bai, M., Sen, B., Wen, S., Ye, H., He, Y., Zhang, X., & Wang, G. (2022). Culturable diversity of Thraustochytrids from coastal waters of Qingdao and their fatty acids. *Marine Drugs*, 20(4), 229.
- Barudin, M. A., Isa, M. L. M., & Yusof, A. M. (2019). Chemical components of polymerase chain reaction in 18s rRNA for detection of *Cryptosporidium* from river water samples. *Malaysian Journal of Analytical Sciences*, 23(3), 401–406.
- Bennett R.M., Honda D., Beakes G.W., Thines M. (2017). *Handbook of the Protists*. Springer; Cham, Switzerland. Labyrinthulomycota; pp. 507–542.
- Bochdansky, A. B., Melissa A. Clouse, & Gerhard J. Herndl. (2017). Eukaryotic microbes, principally fungi and labyrinthulomycetes, dominate biomass on bathypelagic marine snow. *The ISME Journal* 11, no. 2: 362–373.
- Bongiorni L. (2012). Thraustochytrids, a neglected component of organic matter decomposition and food webs in marine sediments. *Progress in molecular and subcellular biology*, 53, 1–13.
- Bongiorni, L., Jain, R., Raghukumar, S., & Aggarwal, R. K. (2005). *Thraustochytrium gaertnerium* sp. nov.: a new thraustochytrid stramenopilan protist from mangroves of Goa, India. *Protist*, 156(3), 303–315.
- Caamaño, E., Loperena, L., Hinzpeter, I., Pradel, P., Gordillo, F., Corsini, G., ... & González, A. R. (2017). Isolation and molecular characterization of

Thraustochytrium strain isolated from Antarctic Peninsula and its biotechnological potential in producing fatty acids. *Brazilian Journal of Microbiology*, 48(4), 671-679.

Cavalier-Smith, T., Allsopp, M. T. E. P., & Chao, E. E. (1994). Thraustochytrids are chromists, not fungi: 18S rRNA signatures of Heterokonta. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 346(1318), 387-397.

Chang, M., Zhang, T., Li, L., Lou, F., Ma, M., Liu, R., ... & Wang, X. (2021). Choreography of multiple omics reveals the mechanism of lipid turnover in *Schizochytrium* sp. S31. *Algal Research*, 54, 102182.

Chauhan, A. S., Patel, A. K., Chen, C. W., Chang, J. S., Michaud, P., Dong, C. Di, & Singhania, R. R. (2023). Enhanced production of high-value polyunsaturated fatty acids (PUFAs) from potential thraustochytrid *Aurantiochytrium* sp. *Bioresource Technology*, 370.

Chauhan, A. S., Patel, A. K., Singhania, R. R., Vadrale, A. P., Chen, C. W., Giri, B. S., ... & Dong, C. D. (2024). Fine-tuning of key parameters to enhance biomass and nutritional polyunsaturated fatty acids production from *Thraustochytrium* sp. *Bioresource technology*, 394, 130252.

Christy, Y. A., Setyati, W. A., & Pribadi, R. (2019). Kajian Valuasi Ekonomi Ekosistem Hutan Mangrove Di Desa Kaliwlingi Dan Desa Sawojajar, Kabupaten Brebes, Jawa Tengah. *Journal of Marine Research*, 8(1), 94-106.

Colonia, B.S.O., de Melo Pereira, G.V., de Carvalho, J.C., dos Santos Sousa, P.H., Fanka, L.S., Rodrigues, C., Medeiros, A.B.P. and Soccol, C.R., 2022. Lipids produced by microalgae and thraustochytrids. In *Biomass, Biofuels, Biochemicals* (pp. 191-217).

Colonia, B.S.O., de Melo Pereira, G.V., Rodrigues, F.M., Muynarsk, E.D.S.M., da Silva Vale, A., de Carvalho, J.C., Soccol, V.T., de Oliveira Penha, R. and Soccol, C.R. 2021. Integrating metagenetics and high-throughput screening for bioprospecting marine thraustochytrids producers of long-chain polyunsaturated fatty acids. *Bioresource Technology*, 333, p.125176.

Damare, V.S., 2019. Advances in isolation and preservation strategies of ecologically important marine protists, the thraustochytrids. In: *Advances in Biological Science Research. Elsevier*, pp. 485–500.

Davidson, R., & Martín del Campo, A. (2020). Combinatorial and computational investigations of neighbor-joining bias. *Frontiers in Genetics*, 11, 584785.

Dellero, Y., Cagnac, O., Rose, S., Seddiki, K., Cussac, M., Morabito, C., Lupette,

- J., Cigliano, R.A., Sanseverino, W., Kuntz, M. and Jouhet, J., 2018. Proposal of a new thraustochytrid genus *Honda* gen. nov. and comparison of its lipid dynamics with the closely related pseudo-cryptic genus *Aurantiochytrium*. *Algal Research*, 35, pp.125-141.
- Douady, C. J., & Nesbø, C. L. (2007). Reconstructing and Interpreting Evolutionary Relationships. *Methods for General and Molecular Microbiology*, 856-868.
- Du, F., Wang, Y. Z., Xu, Y. S., Shi, T. Q., Liu, W. Z., Sun, X. M., & Huang, H. (2021). Biotechnological production of lipid and terpenoid from thraustochytrids. *Biotechnology Advances*, 48, 107725.
- Du, X., Lu, M., Lan, H., Cai, Z., Pan, D., & Wu, Y. (2024). Development of a high-throughput DNA isolation method for livestock and poultry meat based on mesoporous metal-organic framework-coated solid-phase microextraction device. *Journal of Food Composition and Analysis*, 127, 105977.
- Duan, Y., Sen, B., Xie, N., Paterson, J. S., Chen, Z., & Wang, G. (2018). Flow cytometry for rapid enumeration and biomass quantification of thraustochytrids in coastal seawaters. *Microbes and environments*, 33(2), 195-204.
- Dudhagara, D. R., Javia, B. M., & Vala, A. K. (2023). Exploiting marine fungi in the removal of hazardous pollutants and biomass valorisation. In *Marine organisms: a solution to environmental pollution? Uses in bioremediation and in biorefinery* (pp. 117-146). Cham: Springer International Publishing.
- Fan, K. W., Jiang, Y., Faan, Y. W., & Chen, F. (2007). Lipid characterization of mangrove thraustochytrid— *Schizochytrium mangrovei*. *Journal of Agricultural and Food Chemistry*, 55(8), 2906-2910.
- Febrianto, S., Rahman, A., Jati, O. E., Wirasatriya, A., Muskananfolo, M. R., & Latifah, N. (2024). Machine learning for mangrove species distribution using Sentinel 2 satellite image in Segara Anakan, Cilacap Region, Indonesia. *Regional Studies in Marine Science*, 103984.
- Fernández, A., Segura-Alabart, N., & Serratos, F. (2023). The multifurcating neighbor-joining algorithm for reconstructing polytomic phylogenetic trees. *Journal of Molecular Evolution*, 91(6), 773-779.
- Furutani, S., Furutani, N., Kawai, Y., Nakayama, A., & Nagai, H. (2022). Rapid DNA sequencing technology based on the sanger method for bacterial identification. *Sensors*, 22(6), 2130.
- Gil-Gomez, A., Leyland, B., Karthikaichamy, A., Adikes, R. C., Matus, D. Q., Rest, J. S., & Collier, J. L. (2025). Proteome remodeling in the zoospore-to-vegetative cell transition of the stramenopile *Aurantiochytrium limacinum*

- reveals candidate ectoplasmic network proteins. *PloS one*, 20(7), e0326651.
- Ganuza, E., Yang, S., Amezcuita, M., Giraldo-Silva, A., & Andersen, R. A. (2019). Genomics, biology and phylogeny *Aurantiochytrium acetophilum* sp. nov.(Thraustochytriaceae), including first evidence of sexual reproduction. *Protist*, 170(2), 209-232.
- Gupta, A., Singh, D., Byreddy, A. R., Thyagarajan, T., Sonkar, S. P., Mathur, A. S., ... & Puri, M. (2016). Exploring omega-3 fatty acids, enzymes and biodiesel producing thraustochytrids from Australian and Indian marine biodiversity. *Biotechnology journal*, 11(3), 345-355.
- Gupta, A., Wilkens, S., Adcock, J. L., Puri, M., & Barrow, C. J. (2013). Pollen baiting facilitates the isolation of marine thraustochytrids with potential in omega-3 and biodiesel production. *Journal of Industrial Microbiology and Biotechnology*, 40(11), 1231–1240.
- Gupta, A., Worthington, M. J. H., Chalker, J. M., & Puri, M. (2022). Integration of the Exogenous Tuning of Thraustochytrid Fermentation and Sulfur Polymerization of Single-Cell Oil for Developing Plant-like Oils. *Marine Drugs*, 20(10), 1–11.
- Hanrio, E., Severn-Ellis, A., Batley, J., Loh, R., Clode, P., & Dang, C. (2025). A novel thraustochytrid in vitro isolate from the abalone *Haliotis roei* in Western Australia. *Protist*, 126114.
- Honda, D., YOKOCHI, T., NAKAHARA, T., RAGHUKUMAR, S., NAKAGIRI, A., SCHAUMANN, K., & HIGASHIHARA, T. (1999). Molecular phylogeny of labyrinthulids and thraustochytrids based on the sequencing of 18S ribosomal RNA gene. *Journal of Eukaryotic Microbiology*, 46(6), 637-647.
- Huang, J., Liu, Y., Zhu, T., & Yang, Z. (2021). The asymptotic behavior of bootstrap support values in molecular phylogenetics. *Systematic biology*, 70(4), 774-785.
- Istiqomah, M. A., Basyuni, M., & Hasibuan, P. A. Z. (2020). Development Of DNA Extraction on Mangrove Leaves. *International Journal of Science and Technology Research* 9(4), 2277-8616.
- Jape, A., Harsulkar, A., & Sapre, V. R. (2014). Modified Sudan Black B staining method for rapid screening of oleaginous marine yeasts. *International journal of current microbiology and applied sciences*, 3(9), 41-46.
- Kalidasan, K., Asmathunisha, N., Gomathi, V., Dufossé, L., & Kathiresan, K. (2021). Isolation and optimization of culture conditions of *Thraustochytrium kinnei* for biomass production, nanoparticle synthesis,

antioxidant and antimicrobial activities. *Journal of Marine Science and Engineering*, 9(6), 678.

Kalidasan, K., Phusit, H., & Kathiresan, K. (2019). Enumeration of thraustochytrids in decomposing leaves of mangroves as influenced by physicochemical and microbial factors. *J. Curr. Res. Env. App. Myco*, 9, 288-300.

Kalidasan, K., Vinithkumar, N. V., Peter, D. M., Dharani, G., & Dufossé, L. (2021). Thraustochytrids of mangrove habitats from Andaman Islands: Species diversity, PUFA profiles and biotechnological potential. *Marine drugs*, 19(10), 571.

Kaliyamoorthy, K., Kandasamy, K., Chavanich, S., Kamlangdee, N., Vinithkumar, N. V., & Viyakarn, V. (2025). Seasonal dynamics of thraustochytrids in mangrove rhizospheres for microbial interactions, PUFA production. *Scientific Reports*, 15(1), 8027.

Kamlangdee, N., and Fan, K. W. 2003. Polyunsaturated fatty acids production by *Schizochytrium* sp. isolated from mangrove. *Songklanakarinn Journal of Science and Technology*. 25(5): 643-650.

Koopmann, I. K., Müller, B. A., & Labes, A. (2023). Screening of a thraustochytrid strain collection for carotenoid and squalene production characterized by cluster analysis, comparison of 18S rRNA gene sequences, growth behavior, and morphology. *Marine Drugs*, 21(4), 204.

Krock, B. L., Mao, R., Tvrđik, T., Best, D. H., & Lyon, E. (2018). Genomic applications in inherited genetic disorders. In *Genomic Applications in Pathology* (pp. 543-560). Cham: Springer International Publishing.

Lee C., K. J., Dunstan, G. A., Abell, G. C. J., Clementson, L. A., Blackburn, S. I., Nichols, P. D., & Koutoulis, A. (2012). Biodiscovery of new Australian thraustochytrids for production of biodiesel and long-chain omega-3 oils. *Applied Microbiology and Biotechnology*, 93(5), 2215–2231.

Lee, S. H., Erber, W. N., Porwit, A., Tomonaga, M., Peterson, L. C., & International Council for Standardization in Hematology. (2008). ICSH guidelines for the standardization of bone marrow specimens and reports. *International journal of laboratory hematology*, 30(5), 349-364.

Leyland, B., Leu, S., & Boussiba, S. (2017). Are thraustochytrids algae?. *Fungal biology*, 121(10), 835-840.

Leyton, A., Flores, L., Shene, C., Chisti, Y., Larama, G., Asenjo, J. A., & Armenta, R. E. (2021). Antarctic thraustochytrids as sources of carotenoids and high-value fatty acids. *Marine Drugs*, 19(7), 386.

Liu, C., Qi, R. J., Jiang, J. Z., Zhang, M. Q., & Wang, J. Y. (2019). Development

of a blocking primer to inhibit the PCR amplification of the 18S rDNA sequences of *Litopenaeus vannamei* and its efficacy in *Crassostrea hongkongensis*. *Frontiers in microbiology*, *10*, 830.

- Liu, Y., Singh, P., Sun, Y., Luan, S., & Wang, G. (2014). Culturable diversity and biochemical features of thraustochytrids from coastal waters of Southern China. *Applied microbiology and biotechnology*, *98*, 3241-3255.
- Liu, Y., Tang, J., Li, J., Daroch, M., & Cheng, J. J. (2014). Efficient production of triacylglycerols rich in docosahexaenoic acid (DHA) by osmo-heterotrophic marine protists. *Applied microbiology and Biotechnology*, *98*(23), 9643-9652.
- Lyu, L., Wang, Q., & Wang, G. (2021). Cultivation and diversity analysis of novel marine thraustochytrids. *Marine Life Science & Technology*, *3*, 263-275.
- Manikan, V., Nazir, M. Y. M., Kalil, M. S., Isa, M. H. M., Kader, A. J. A., Yusoff, W. M. W., & Hamid, A. A. (2015). A new strain of docosahexaenoic acid producing microalga from Malaysian coastal waters. *Algal Research*, *9*, 40-47.
- Marchan, L. F., Chang, K. J. L., Nichols, P. D., Mitchell, W. J., Polglase, J. L., & Gutierrez, T. (2018). Taxonomy, ecology and biotechnological applications of thraustochytrids: A review. *Biotechnology advances*, *36*(1), 26-46.
- Matsubara, T., Soh, J., Morita, M., Uwabo, T., Tomida, S., Fujiwara, T., Kanazawa, S., Toyooka, S. and Hirasawa, A., (2020). DV200 index for assessing RNA integrity in next-generation sequencing. *BioMed research international*, *2020*(1), p.9349132.
- Menzorov, A. G., Iukhtanov, D. A., Naumenko, L. G., Bobrovskikh, A. V., Zubairova, U. S., Morozova, K. N., & Doroshkov, A. V. (2024). Thraustochytrids: Evolution, ultrastructure, biotechnology, and modeling. *International Journal of Molecular Sciences*, *25*(23), 13172.
- Mo, C., Douek, J., & Rinkevich, B. (2002). Development of a PCR strategy for thraustochytrid identification based on 18S rDNA sequence. *Marine Biology*, *140*, 883–889.
- Morabito, C., Bournaud, C., Maës, C., Schuler, M., Cigliano, R. A., Dellerio, Y., Eric, M., Alberto, A., & Rébeillé, F. (2019). The lipid metabolism in thraustochytrids. *Progress in Lipid Research*, *76*, 101007.
- Nakai, R., Nakamura, K., Jadoon, W. A., Kashihara, K., & Naganuma, T. (2013). Genus-specific quantitative PCR of thraustochytrid protists. *Marine Ecology Progress Series*, *486*, 1-12.
- Nakazawa, A., Kokubun, Y., Matsuura, H., Yonezawa, N., Kose, R., Yoshida, M.,

- Tanabe, Y., Kusuda, E., Van Thang, D., Ueda, M. and Honda, D., 2014. TLC screening of thraustochytrid strains for squalene production. *Journal of Applied Phycology*, 26(1), pp.29-41.
- Nasution, M. Y. A., & Wijaya, H. B. (2024). Pengelolaan Kawasan Mangrove Berbasis Masyarakat di Kecamatan Sayung, Demak. *Teknik PWK (Perencanaan Wilayah Kota)*, 13(3), 189-196.
- Park, H., Kwak, M., Seo, J., Ju, J., Heo, S., Park, S., & Hong, W. (2018). Enhanced production of carotenoids using a Thraustochytrid microalgal strain containing high levels of docosahexaenoic acid-rich oil. *Bioprocess and Biosystems Engineering*, 41(9), 1355-1370.
- Phumphumirat, W., Ferguson, D. K., & Gleason, F. H. (2016). The colonization of palynomorphs by chytrids and thraustochytrids during pre-depositional taphonomic processes in tropical mangrove ecosystems. *Fungal ecology*, 23, 11-19.
- Qarri, A., Rinkevich, Y., & Rinkevich, B. (2021). Employing marine invertebrate cell culture media for isolation and cultivation of thraustochytrids. *Botanica Marina*, 64(6), 447-454.
- Qarri, A., Rinkevich, Y., Douek, J., Sardogan, A., & Rinkevich, B. (2024). Growth Performance of a Newly Isolated and Culturable Thraustochytrid Strain from Sea Squirt Colonies. *Fishes*, 9(1), 22.
- Quilodrán, B., Cortinez, G., Bravo, A., & Silva, D. (2020). Characterization and comparison of lipid and PUFA production by native thraustochytrid strains using complex carbon sources. *Heliyon*, 6(11).
- Raghukumar, S., & Damare V.S. 2011. Increasing evidence for the important role of labyrinthulomycetes in marine ecosystems. *Botanica Marina*; 54:3-11.
- Rahim, A., Soeprbowati, T. R., & Putranto, T. T. (2023). Mangrove species structure at the Tuntang Estuary, Demak, Central Java. In *IOP Conference Series: Earth and Environmental Science* (Vol. 1137, No. 1, p. 012053). IOP Publishing.
- Ramirez, J. L., Santos, C. A., Machado, C. B., Oliveira, A. K., Garavello, J. C., Britski, H. A., & Galetti Jr, P. M. (2020). Molecular phylogeny and species delimitation of the genus *Schizodon* (Characiformes, Anostomidae). *Molecular phylogenetics and evolution*, 153, 106959.
- Ren, L. J., Sun, G. N., Ji, X. J., Hu, X. C., & Huang, H. (2014). Compositional shift in lipid fractions during lipid accumulation and turnover in *Schizochytrium* sp. *Bioresource technology*, 157, 107-113.
- Rollin, S., Gupta, A., & Puri, M. (2022). Optimising pineapple filtrate assisted cell

- disruption of wet thraustochytrid biomass for improved lipid extraction. *Journal of Cleaner Production*, 378, 134393.
- Rosa, S. M., Galvagno, M. A., & Vélez, C. G. (2011). Adjusting culture conditions to isolate thraustochytrids from temperate and cold environments in southern Argentina. *Mycoscience*, 52(4), 242-252.
- Rozas, J., Ferrer-Mata, A., Sánchez-DelBarrio, J. C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S. E., & Sánchez-Gracia, A. (2017). DnaSP 6: DNA sequence polymorphism analysis of large datasets. *Molecular Biology and Evolution*, 34, 10.1093/molbev/msx248.
- Sari, S. P. W., & Rifai, A. (2020). Pengelolaan Desa Wisata Hutan Mangrove Desa Bedono Kecamatan Sayung Kabupaten Demak. *Aksara: Jurnal Ilmu Pendidikan Nonformal*, 6(2), 121-138.
- Shakeri, S., Amoozyan, N., Fekrat, F., & Maleki, M. (2017). Antigastric cancer bioactive Aurantiochytrium oil rich in docosahexaenoic acid: from media optimization to cancer cells cytotoxicity assessment. *Journal of food science*, 82(11), 2706-2718.
- Sik Kim, H., Hyun Byun, S., & Mu Lee, B. (2005). Effects of chemical carcinogens and physicochemical factors on the UV spectrophotometric determination of DNA. *Journal of Toxicology and Environmental Health, Part A*, 68(23-24), 2081-2095.
- Siriraka, K., Suanjitb, S., Powtongsookc, S., & Jaritkhuane, S. (2020). Characterization and PUFA production of Aurantiochytrium limacinum BUCHAXM 122 isolated from fallen mangrove leaves. *ScienceAsia*, 46(4), 403-411.
- Song, Y., Yang, X., Li, S., Luo, Y., Chang, J.-S., & Hu, Z. (2023). Thraustochytrids as a promising source of fatty acids, carotenoids, and sterols: Bioactive compound biosynthesis, and modern biotechnology. *Critical Reviews in Biotechnology*, 44, 1-23.
- Stokes, N. A., Ragone Calvo, L. M., Reece, K. S., & Burreson, E. M. (2002). Molecular diagnostics, field validation, and phylogenetic analysis of Quahog Parasite Unknown (QPX), a pathogen of the hard clam *Mercenaria mercenaria*. *Diseases of Aquatic Organisms*, 52, 233–247.
- Subagiyo, S., Djarod, M. S. R., & Setyati, W. A. (2017). Potensi Ekosistem Mangrove Sebagai Sumber Bakteri Untuk Produksi Protease, Amilase Dan Selulase. *Jurnal Kelautan Tropis*, 20(2), 106-111.
- Suhendra, S., Pantoiyo, T., Fazlia, S., Sulistiawati, E., & Evitasari, R. T. (2021). Bioprocess Potentials of Squalene from Thraustochytrids Microalgae for

Nutraceuticals in New Normal Era Isolated from Indonesian Mangroves: A Review. *CHEMICA. Jurnal Teknik Kimia*, 8(1), 18.

- Suhendra, Zahro, H., Sulistiawati, E., Neubauer, P., & Hutari, A. (2019). Kajian Singkat Potensi Rancang Bangun Pabrik Omega-3 (DHA) Kemurnian Tinggi Berbahan Baku Spesies *Aurantiochytrium* dari Hutan Bakau Indonesia untuk Menunjang Ketahanan Pangan Nasional. *Konversi*, 8(1), 33–44.
- Tamura, K., Stecher, G., & Kumar, S. (2021). MEGA11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology and Evolution*, 38(7), 3022–3027.
- Tsui, C. K., KW, F., Chow, R. K., EB, G. J., & Vrijmoed, L. L. (2012). Zoospore production and motility of mangrove thraustochytrids from Hong Kong under various salinities. *Mycoscience*, 53(1), 1-9.
- Ueda, M., Nomura, Y., Doi, K., Nakajima, M., & Honda, D. (2015). Seasonal dynamics of culturable thraustochytrids (Labyrinthulomycetes, Stramenopiles) in estuarine and coastal waters. *Aquatic Microbial Ecology*, 74(3), 187-204.
- Ulken, A. (1983). Distribution of Phycomycetes in mangrove swamps with brackish waters and waters of high salinity. In *Biology and ecology of mangroves* (pp. 111-116). Dordrecht: Springer Netherlands. Ulken, A. (1983). Distribution of Phycomycetes in mangrove swamps with brackish waters and waters of high salinity. In *Biology and ecology of mangroves* (pp. 111-116). Dordrecht: Springer Netherlands.
- Wang, Q., Han, W., Jin, W., Gao, S., & Zhou, X. (2021). Docosaheanoic acid production by *Schizochytrium* sp.: Review and prospect. *Food Biotechnology*, 35(2), 111-135.
- Wang, Q., Ye, H., Xie, Y., He, Y., Sen, B., & Wang, G. (2019). Culturable diversity and lipid production profile of Labyrinthulomycete protists isolated from coastal mangrove habitats of China. *Marine drugs*, 17(5), 268.